

# Results of active surveillance in wild birds for HPAI in Italy



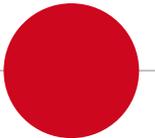
**Federica Gobbo, DVM**

*EU Reference Laboratory for Avian Influenza  
and Newcastle Disease*



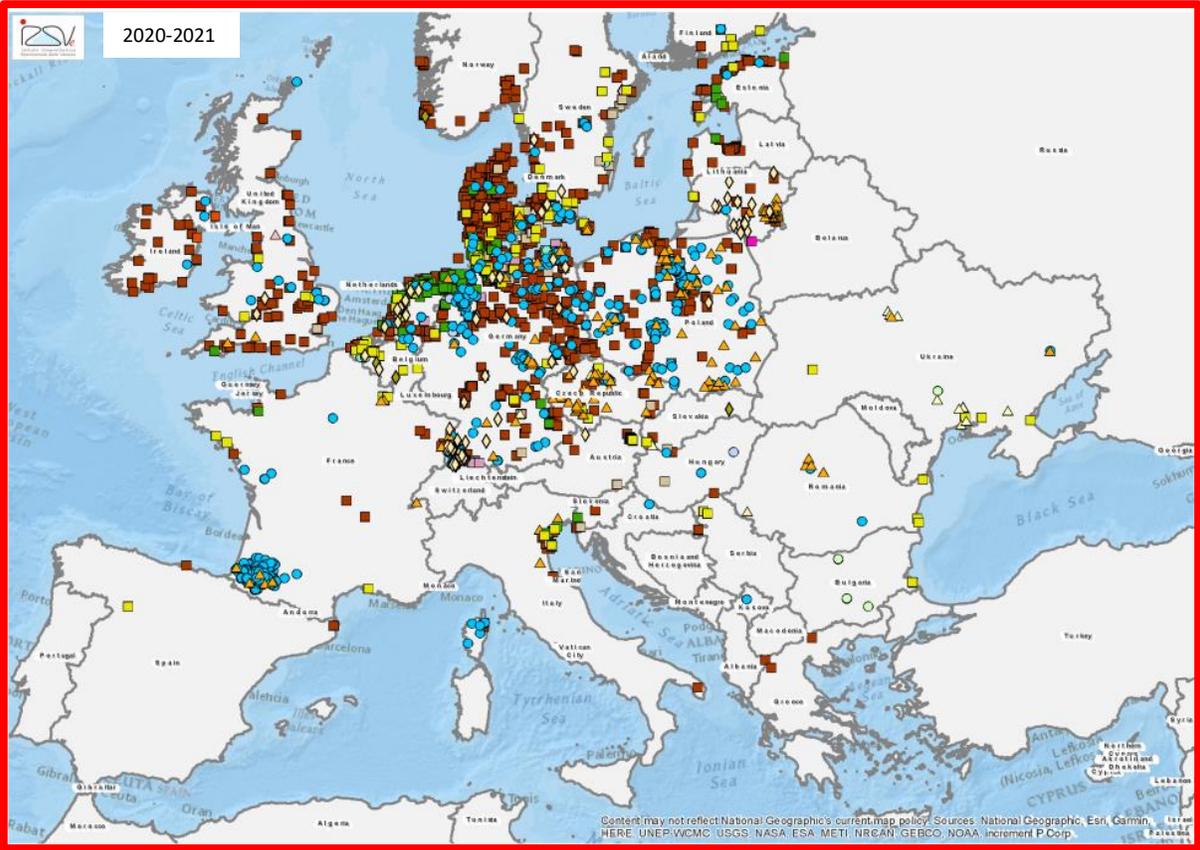
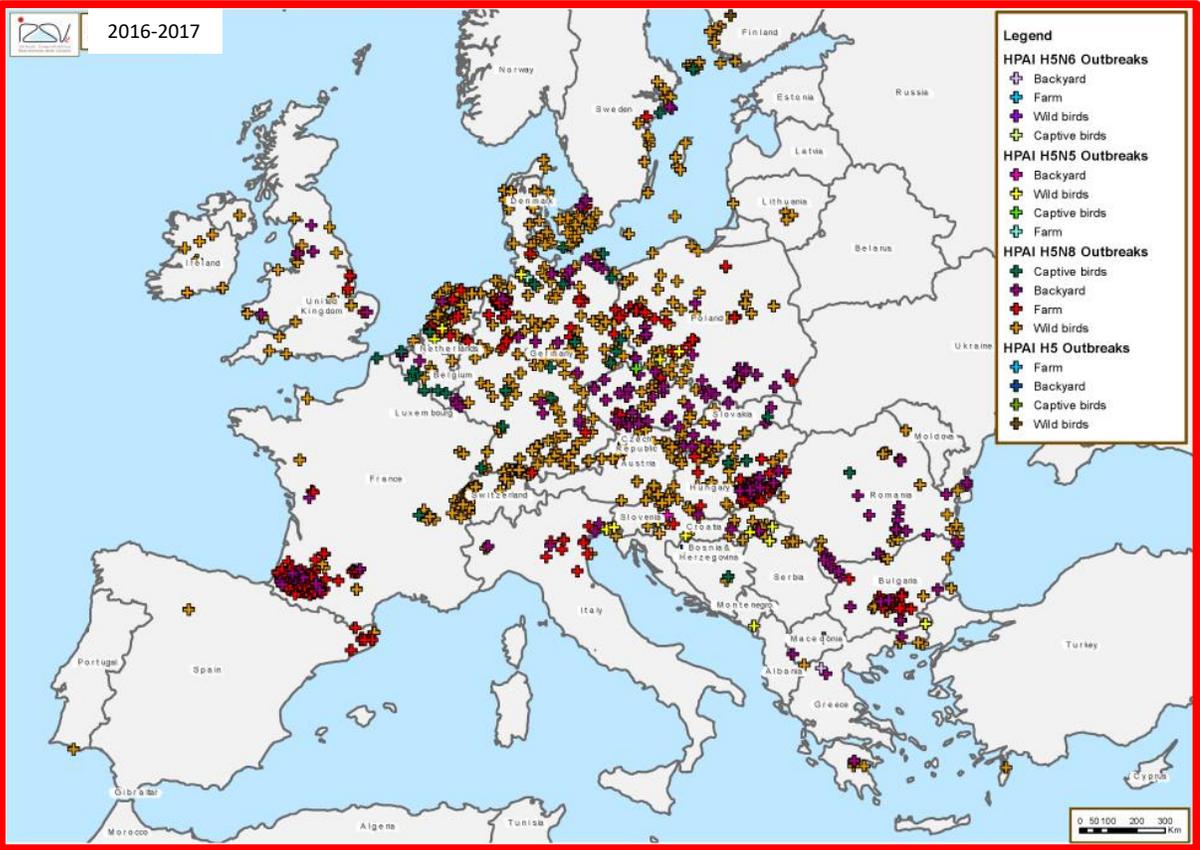
**27<sup>th</sup> Annual Meeting**

of the National Reference Laboratories for Avian Influenza  
and Newcastle Disease of European Union Member States



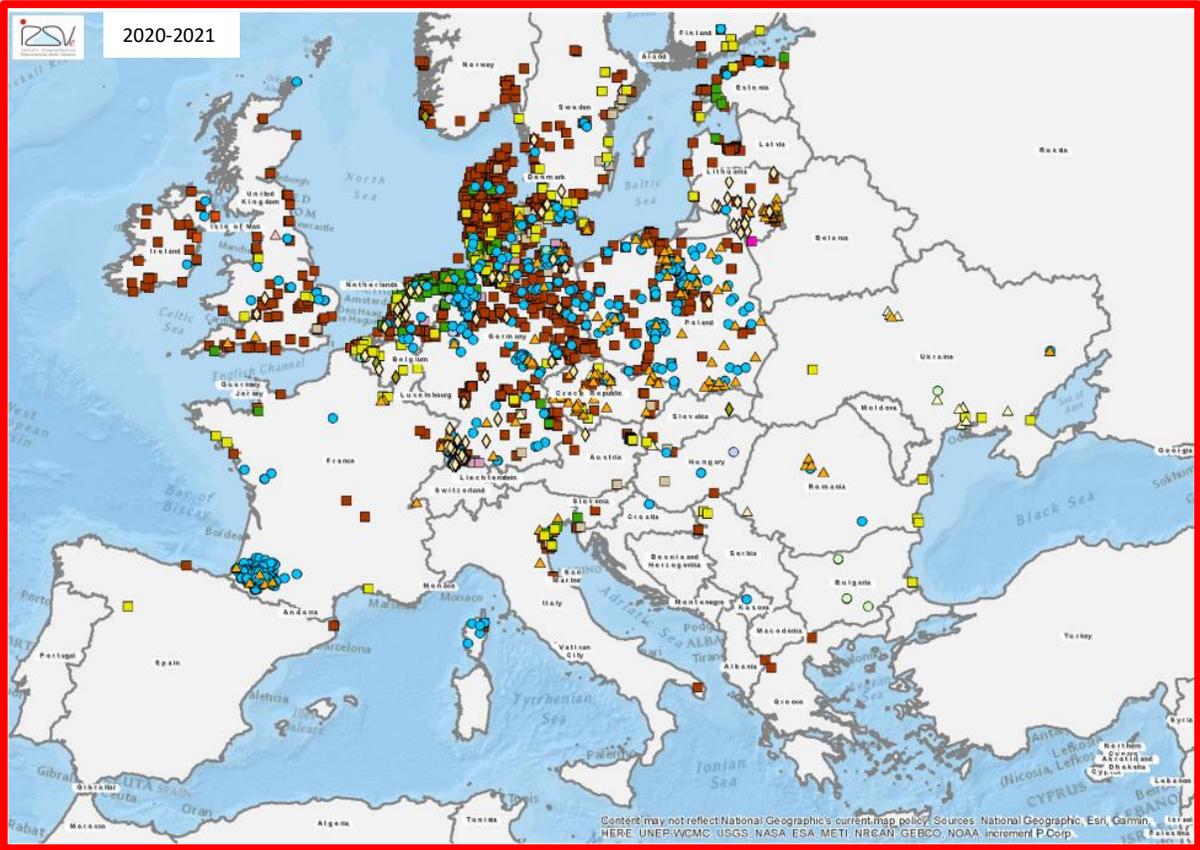
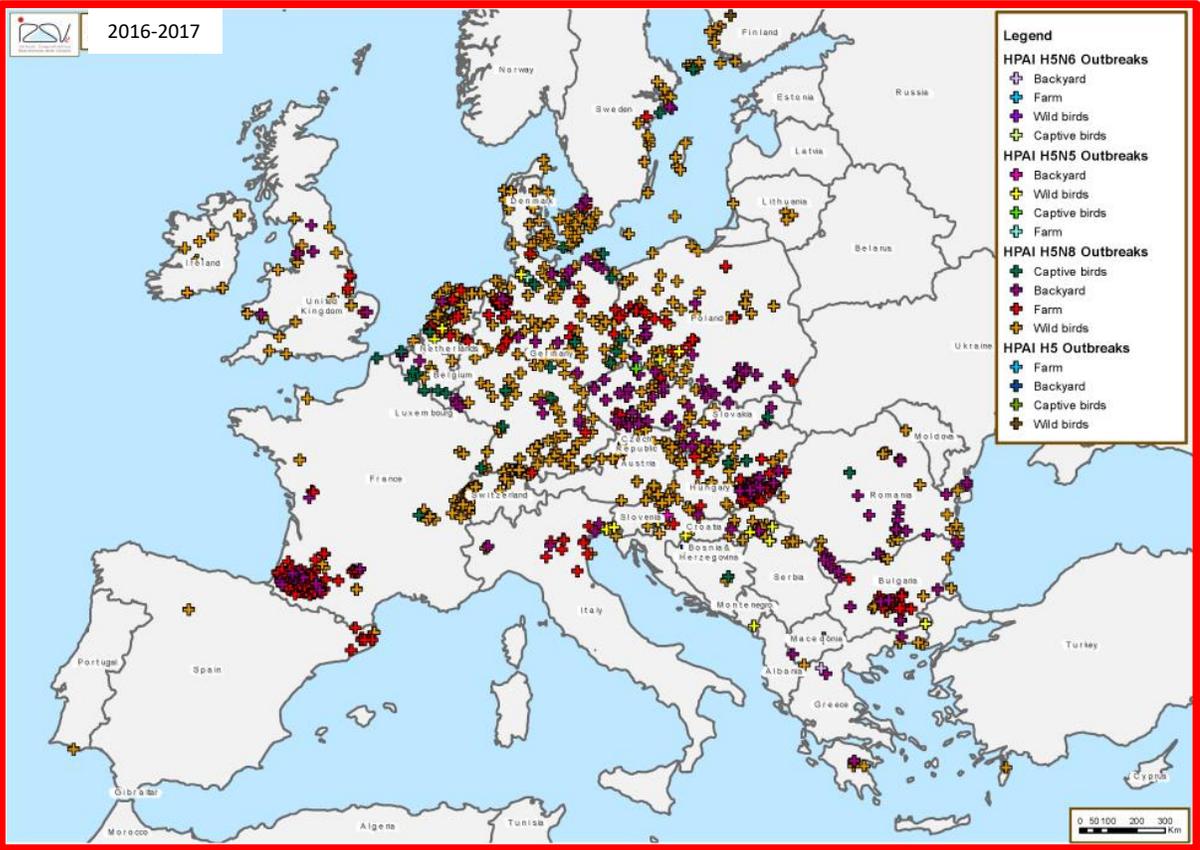
# Background

HPAI H5 viruses belonging to clade 2.3.4.4b of the Goose/Guangdong/96 (GS/Gd) lineage



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# The Italian perspective

**Decree of the Italian Ministry of Health of 14 March 2018:** ancillary activities were implemented to enhance the AI National Surveillance Plan



Pilot surveillance activities in wetlands of the northern Italy



Strategic zone for the introduction of AI *via* migratory flows

**Early detection**

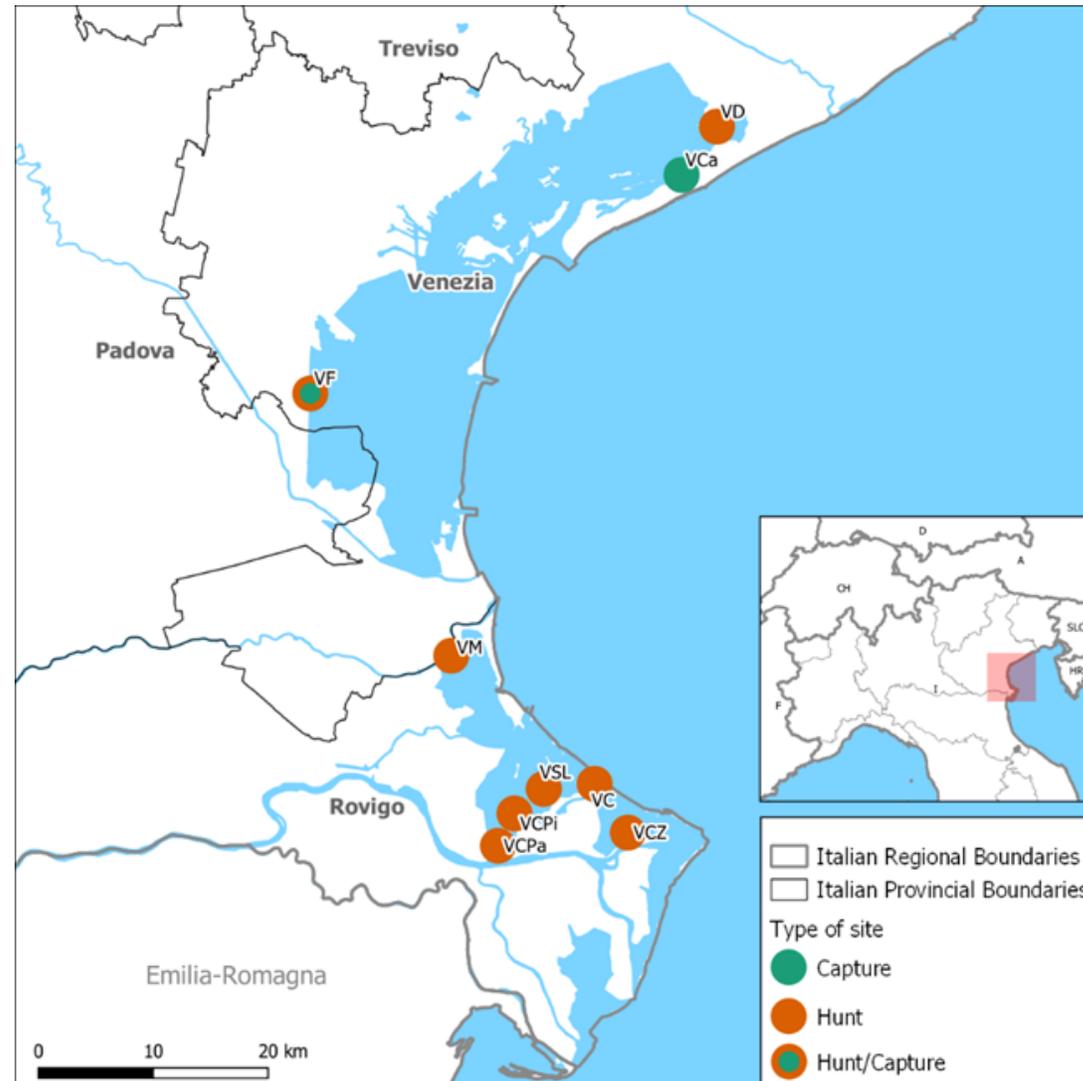


Wild-domestic bird interface: unique combination of densely populated poultry areas (DPPAs) and wetlands

**Risk-based**

1. Active Surveillance in hunted dabbling ducks (Nov 2020-Jan 2021)

2. Active Surveillance in birds Capture Station (Nov 2020-March 2021)



Location of hunting sites (orange color) and capture sites (green color). V, Valle; **VCPa**, V. Ca Pasta (RO); **VCZ**, V. Ca Zuliani (RO); **VD**, V. Drago (VE); **VSL**, V. San Leonardo (RO); **VC**, V. Chiusa (RO); **VCPi**, V. Ca Pisani (RO); **VM**, V. Morosina (RO); **VF**, Valle Figheri (VE); **VCa**, Valle Cavallino.

## Hunted dabbling ducks

- 8 wetlands selected for the active surveillance (Province of Rovigo and Venice)
- 15 samplings from November 2020 to January 2021 (a total of **823 hunted** aquatic birds were sampled via oropharyngeal swab (**OS**), cloacal swab (**CS**) and feather swab (**FS**))

337 EW

220 ET

111 Ma

49 Ga

94 NS

12 NP



*Eurasian wigeon*

*Eurasian teal*

*Mallard*

*Gadwall*

*Northern shoveler*

*Northern pintail*

**2449** animal swabs were investigated for AIV (M-gene, H5,H7,H9). The identified viruses were characterized and pathotyped by sequencing. AI viruses were easily isolated in SPF chicken eggs from selected samples

Molecular detection and prevalence of AIVs in different biological samples collected from **hunted dabbling ducks** (Veneto region, Northern Italy, 2020-2021). Blue, HPAIV circulation; yellow, LPAIV circulation; green, HPAIV and LPAIV co-circulation.

HVSS ID	Sampling yy-mm-dd	Eurasian Teal			Gadwall			Northern Pintail			Eurasian Wigeon			Mallard			Northern Shoveler			Dabbling ducks									
		ET	Pos. N. (%)			Ga	Pos. N. (%)			NP	Pos. N. (%)			EW	Pos. N. (%)			Ma	Pos. N. (%)			NS	Pos. N. (%)			DD	Pos. N. (%)		
			OS	CS	FS		OS	CS	FS		OS	CS	FS		OS	CS	FS		OS	CS	FS		OS	CS	FS		OS	CS	FS
VCPa-1 <sup>st</sup>	20-11-09	7	0	1 (14.3)	0	19	0	0	0	1	0	0	0	6	0	2 (33.3)	0	8	0	0	0	13	0	0	0	54	0	3 (5.6)	0
VCZ-1 <sup>st</sup>	20-11-16	1	0	0	0	5	0	0	1 (20)	4	0	0	0	10	1 (10)	2 (20)	0	19	1 (5.3)	1 (5.3)	2 (10.5)	1	0	0	0	40	2 (5)	3 (7.5)	3 (7.5)
VD-1 <sup>st</sup>	20-11-21	0	—	—	—	4	0	0	0	5	0	0	0	100	10 (10)	9 (9)	5 (5)	0	—	—	—	11	0	1 (9.1)	1 (9.1)	120	10 (8.3)	10 (8.3)	6 (5)
VSL-1 <sup>st</sup>	20-11-21	8	0	0	0	0	—	—	—	0	—	—	—	54	2 (3.7)	3 (5.6)	1 (1.9)	8	0	0	0	40	0	0	0	110	2 (1.8)	3 (2.7)	1 (0.9)
VC-1 <sup>st</sup>	20-11-26	32	2 (6.3)	0	2 (6.3)	2	0	0	0	0	—	—	—	16	0	0	2 (12.5)	8	0	0	1 (12.5)	4	0	0	0	62	2 (3.2)	0	5 (8.1)
VD-2 <sup>nd</sup>	20-11-28	0	—	—	—	0	—	—	—	0	—	—	—	20	0	0	—	0	—	—	—	0	—	—	—	20	0	0	—
VCPI	20-12-07	46	1 (2.2)	0	0	0	—	—	—	0	—	—	—	2	0	0	0	2	0	0	0	0	—	—	—	50	1 (2)	0	0
VCPa-2 <sup>nd</sup>	20-12-15	1	0	0	0	3	0	0	0	0	—	—	—	89	3 (3.4)	4 (4.5)	1 (1.1)	5	0	0	0	2	0	0	0	100	3 (3)	4 (4)	1 (1)
VM-1 <sup>st</sup>	20-12-22	60	1 (1.7)	0	0	2	0	0	0	0	—	—	—	0	—	—	—	23	0	0	0	1	0	0	0	86	1 (1.2)	0	0
VC-2 <sup>nd</sup>	21-01-09	20	0	0	0	1	0	0	0	0	—	—	—	16	0	0	0	4	0	0	0	4	0	0	0	45	0	0	0
VD-3 <sup>rd</sup>	21-01-11	15	0	1 (6.7)	0	0	—	—	—	0	—	—	—	15	0	0	0	0	—	—	—	0	—	—	—	30	0	1 (3.3)	0
VSL-2 <sup>nd</sup>	21-01-16	1	0	0	0	0	—	—	—	0	—	—	—	4	0	0	0	6	0	0	0	7	0	1 (14.3)	0	18	0	1 (5.6)	0
VF	21-01-24	1	0	0	0	11	0	0	0	2	0	0	0	2	0	0	0	15	0	0	0	8	0	1 (12.5)	0	39	0	1 (2.6)	0
VCZ-2 <sup>nd</sup>	21-01-25	3	1 (33.3)	0	0	0	—	—	—	0	—	—	—	3	1 (33.3)	0	2 (66.7)	5	1 (20)	0	1 (20)	2	1 (50)	0	0	13	4 (30.8)	0	3 (23.1)
VM-2 <sup>nd</sup>	21-01-30	25	0	1 (4)	0	2	0	0	0	0	—	—	—	0	—	—	—	8	0	0	0	1	0	0	0	36	0	1 (2.8)	0
<b>Total duck &amp; Pos. (%)</b>	20-11-09 21- 01-30	220	5 (2.3)	3 (1.4)	2 (0.9)	49	0	0	1 (2)	12	0	0	0	337	17 (5)	20 (5.9)	11 (3.3)	111	2 (1.8)	1 (0.9)	4 (3.6)	94	1 (1.1)	3 (3.2)	1 (1.1)	823	25 (3)	27 (3.3)	19 (2.3)



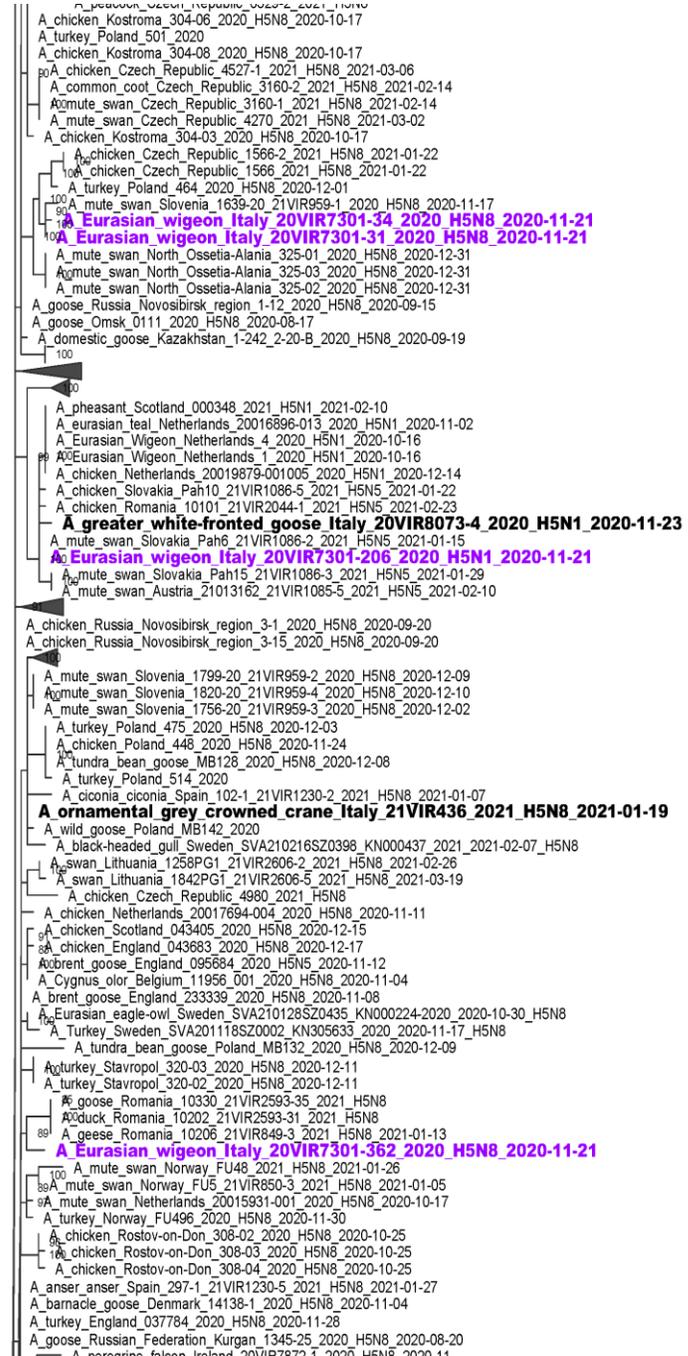


HA

- HPAI H5 viruses, Valle Ca' Zuliani, Rovigo
- HPAI H5 viruses, Valle Chiusa, Rovigo
- HPAI H5 viruses, Valle Drago, Venice
- HPAI H5 viruses, Valle Figheri, Venice



Clade 2.3.4.B



Clade 2.3.4.B

## Captured dabbling ducks

- 2 wetlands selected for active surveillance in Province of Venice: Valle Figheri wetland (14 samplings from November 2020 to March 2021) and Valle Cavallino wetland (2 samplings in February 2021)
- **521** captured aquatic birds were sampled via oropharyngeal swab (**OS**), cloacal swab (**CS**) and feather swab (**FS**)

4 EW



*Eurasian wigeon*

374 ET



*Eurasian teal*

61 Ma



*Mallard*

2 Ga



*Gadwall*

1 CSh



*Common shelduck*

79 NP



*Northern pintail*

**1563** animal swabs were investigated for AIV (M-gene, H5,H7,H9). The identified viruses were characterized and pathotyped by sequencing. AI viruses were easily isolated in SPF chicken eggs from selected samples

Molecular detection and prevalence of AIVs in different biological samples collected from **captured dabbling ducks** (Veneto region, Northern Italy, 2020-2021). Blue, HPAIV circulation; yellow, LPAIV circulation; green, HPAIV and LPAIV co-circulation.

TSS ID	Sampling yy-mm-dd	ET	Eurasian Teal			Gadwall			Northern Pintail			Eurasian Wigeon			Mallard			Common Shelduck			Dabbling ducks								
			Pos. N. (%)			Ga	Pos. N. (%)			NP	Pos. N. (%)			EW	Pos. N. (%)			Ma	Pos. N. (%)			CSh	Pos. N. (%)			DD	Pos. N. (%)		
			OS	CS	FS		OS	CS	FS		OS	CS	FS		OS	CS	FS		OS	CS	FS		OS	CS	FS		OS	CS	FS
VF-1	20-11-12	24	1 (4.2)	0	0	0	—	—	—	0	—	—	—	0	—	—	—	3	0	0	0	0	—	—	—	27	1 (3.7)	0	0
VF-2	20-11-12	10	0	1 (10)	0	0	—	—	—	0	—	—	—	0	—	—	—	3	0	0	0	0	—	—	—	13	0	1 (7.7)	0
VF-3	20-11-27	24	0	2 (8.3)	0	0	—	—	—	0	—	—	—	0	—	—	—	2	0	0	0	0	—	—	—	26	0	2 (7.7)	0
VF-4	20-12-04	68	15 (22.1)	4 (5.9)	9 (13.2)	0	—	—	—	0	—	—	—	0	—	—	—	1	0	0	0	0	—	—	—	69	15 (21.7)	4 (5.8)	9 (13)
VF-5	20-12-11	36	7 (19.4)	0	1 (2.8)	0	—	—	—	0	—	—	—	0	—	—	—	2	0	0	0	0	—	—	—	38	7 (18.4)	0	1 (2.6)
VF-6	20-12-18	24	2 (8.3)	1 (4.2)	0	0	—	—	—	0	—	—	—	0	—	—	—	2	0	0	0	0	—	—	—	26	2 (7.7)	1 (3.8)	0
VF-7	20-12-23	6	0	0	0	0	—	—	—	0	—	—	—	0	—	—	—	1	0	0	0	0	—	—	—	7	0	0	0
VF-8	21-01-07	10	0	1 (10)	0	0	—	—	—	0	—	—	—	0	—	—	—	1*	0	0	0	0	—	—	—	11	0	1 (9.1)	0
VF-9	21-01-14	6	0	0	0	0	—	—	—	0	—	—	—	0	—	—	—	10	0	0	1 (10)	0	—	—	—	16	0	0	1 (6.3)
VF-10	21-01-22	40	0	2 (5)	0	0	—	—	—	0	—	—	—	0	—	—	—	6**	0	0	0	0	—	—	—	46	0	2 (4.3)	0
VF-11	21-01-29	51	1 (2)	3 (5.9)	0	2	0	0	0	0	—	—	—	0	—	—	—	9	1 (11.1)	0	0	0	—	—	—	62	2 (3.2)	3 (4.8)	0
VF-12	21-02-22	19	0	0	0	0	—	—	—	0	—	—	—	0	—	—	—	2	0	0	0	0	—	—	—	21	0	0	0
VF-13	21-02-25	29	0	2 (6.9)	0	0	—	—	—	0	—	—	—	0	—	—	—	5	0	0	0	0	—	—	—	34	0	2 (5.9)	0
VF-14	21-03-05	9	0	0	0	0	—	—	—	0	—	—	—	0	—	—	—	14	0	0	0	0	—	—	—	23	0	0	0
VCa-1	21-02-12	18	0	0	0	0	—	—	—	74	0	3 (4.1)	0	4	0	0	0	0	—	—	—	1	0	1 (100)	0	97	0	4 (4.1)	0
VCa-2	21-02-15	0	—	—	—	0	—	—	—	5	0	1 (20)	0	0	—	—	—	0	—	—	—	0	—	—	—	5	0	1 (20)	0
<b>Total duck &amp; Pos. (%)</b>	20-11-12 21-03-05	374	26 (7)	16 (4.3)	10 (2.7)	2	0	0	0	79	0	4 (5.1)	0	4	0	0	0	61	1 (1.6)	0	1 (1.6)	1	0	1 (100)	0	521	27 (5.2)	21 (4)	11 (2.1)

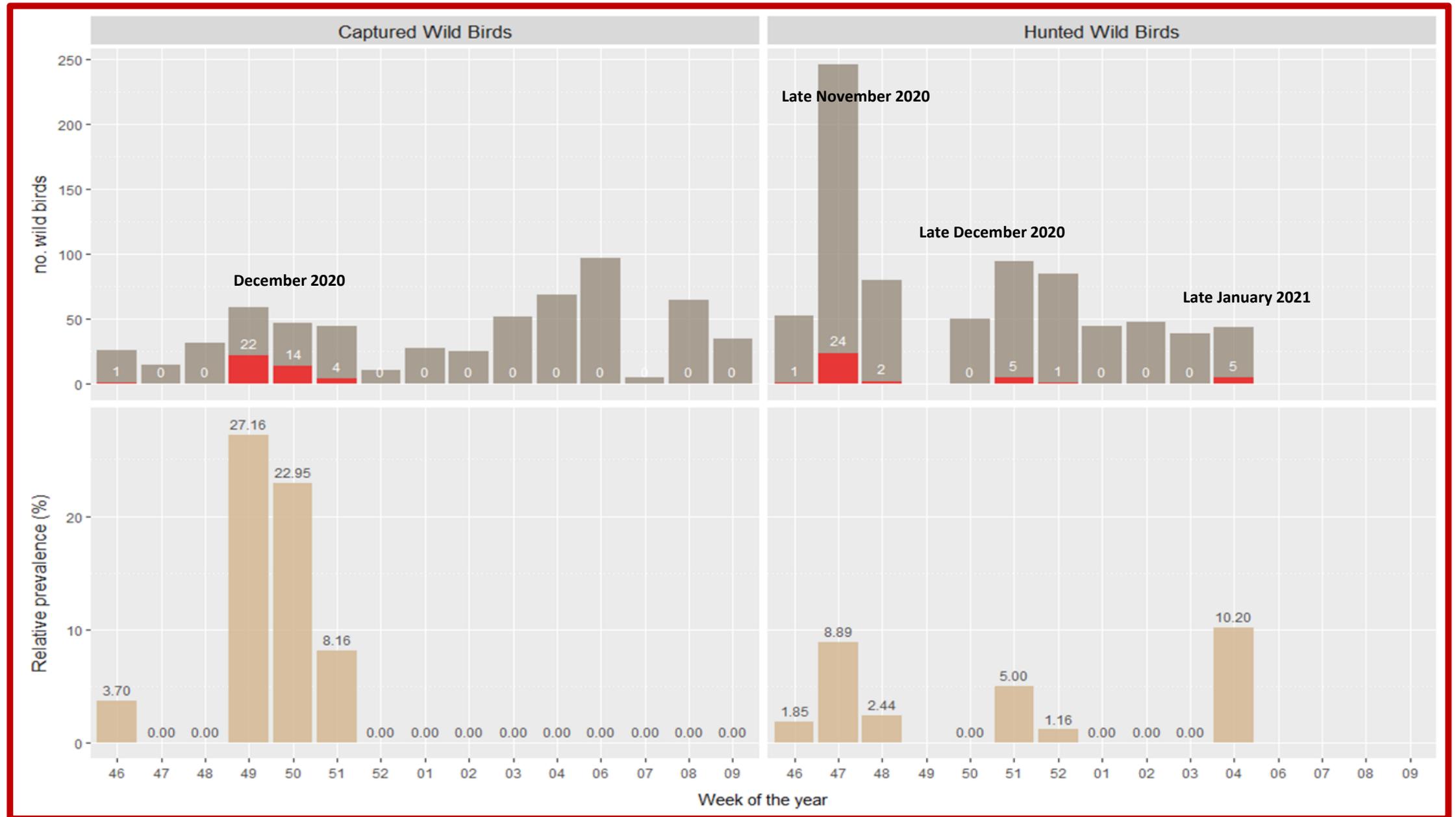
Molecular pathotyping and subtyping of AIVs detected in different biological samples collected from captured dabbling ducks  
(Veneto region, Northern Italy, 2020-2021).

TSS ID (yy-mm-dd)	AIV Pos		OS virological results				CS virological results				FS virological results						
	C-duck ID	Sex	Age	AIV	M	AIV	P-T	AIV	M	AIV	P-T	AIV	M	AIV	P-T	AIV	S-
				gene		gene		gene		gene		gene		gene		gene	
VF-1 (20-11-12)	C1_ET	M	J	Pos.		HPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.				
VF-2 (20-11-20)	C2_ET	F	A	-	n.d.	n.d.		Pos.	LPAL	H5Nx	-	n.d.	n.d.				
VF-3 (20-11-27)	C3_ET	M	J	-	n.d.	n.d.		Pos.	LPAL	HxNx	-	n.d.	n.d.				
	C4_ET	M	J	-	n.d.	n.d.		Pos.	LPAL	HxNx	-	n.d.	n.d.				
VF-4 (20-12-04)	C5_ET	F	J	Pos.	HPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.					
	C6_ET	M	J	Pos.	HPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.					
	C7_ET	F	J	Pos.	HPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.					
	C8_ET	F	J	Pos.	HPAI	H5Nx	Pos.	HPAI	H5Nx	-	n.d.	n.d.					
	C9_ET	F	J	Pos.	HPAI	H5Nx	-	n.d.	n.d.	Pos.	HPAI	H5Nx					
	C10_ET	F	A	Pos.	LPAL	HxNx	-	n.d.	n.d.	-	n.d.	n.d.					
	C11_ET	F	A	Pos.	HPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.					
	C12_ET	M	J	Pos.	HPAI	H5N8	Pos.	HPAI	H5Nx	-	n.d.	n.d.					
	C13_ET	M	A	Pos.	HPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.					
	C14_ET	F	J	Pos.	HPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.					
	C15_ET	M	J	Pos.	HPAI	H5N8	Pos.	HPAI	H5Nx	-	n.d.	n.d.					
	C16_ET	F	J	-	n.d.	n.d.	Pos.	LPAL	HxNx	-	n.d.	n.d.					
	C17_ET	F	J	Pos.	HPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.					
	C18_ET	F	J	Pos.	HPAI	H5Nx	-	n.d.	n.d.	Pos.	HPAI	H5Nx					
	C19_ET	F	J	Pos.	HPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.					
	C20_ET	M	A	Pos.	HPAI	H5N1	-	n.d.	n.d.	Pos.	HPAI	H5Nx					
C21_ET	M	J	-	n.d.	n.d.	-	n.d.	n.d.	Pos.	HPAI	H5Nx						
C22_ET	F	J	-	n.d.	n.d.	-	n.d.	n.d.	Pos.	HPAI	H5Nx						
C23_ET	M	J	-	n.d.	n.d.	-	n.d.	n.d.	Pos.	HPAI	H5Nx						
C24_ET	F	A	-	n.d.	n.d.	-	n.d.	n.d.	Pos.	HPAI	H5Nx						
C25_ET	F	J	-	n.d.	n.d.	-	n.d.	n.d.	Pos.	HPAI	H5Nx						
C26_ET	F	J	-	n.d.	n.d.	-	n.d.	n.d.	Pos.	HPAI	H5Nx						
C27_ET	M	J	Pos.	HPAI	H5N8	-	n.d.	n.d.	-	n.d.	n.d.						
C28_ET	F	J	Pos.	HPAI	H5N8	-	n.d.	n.d.	-	n.d.	n.d.						
C29_ET	F	J	Pos.	HPAI	H5N8	-	n.d.	n.d.	-	n.d.	n.d.						
VF-5 (20-12-11)	C30_ET	F	J	Pos.	HPAI	H5N1-N8	-	n.d.	n.d.	-	n.d.	n.d.					
	C31_ET	F	A	Pos.	HPAI	H5N8	-	n.d.	n.d.	-	n.d.	n.d.					
	C32_ET	F	J	Pos.	HPAI	H5N8	-	n.d.	n.d.	-	n.d.	n.d.					
	C33_ET	F	J	Pos.	n.a.	H5N8	-	n.d.	n.d.	-	n.d.	n.d.					
	C34_ET	F	J	-	n.d.	n.d.	-	n.d.	n.d.	Pos.	HPAI	H5N8					

Cont.

TSS ID (yy-mm-dd)	AIV Pos.			OS virological results			CS virological results					FS virological results							
	C-duck ID	Sex	Age	AIV	M	AIV	P-T	AIV	M	AIV	P-T	AIV	S-T	AIV	M	AIV	P-T	AIV	S-
				gene	gene	gene	gene	gene	gene	gene	gene	gene	gene	gene	gene	gene	gene	gene	gene
	C35_ET	F	J	Pos.	HPAI	H5N8	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
VF-6 (20-12-18)	C36_ET	M	J	Pos.	HPAI	H5N8	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C37_ET	F	J	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
VF-8 (21-01-07)	C38_ET	F	J	-	n.d.	n.d.	Pos.	LPAI	H7N3	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
VF-9 (21-01-14)	C39_Ma	F	J	-	n.d.	n.d.	-	n.d.	n.d.	Pos.	LPAI	H9Nx	-	n.d.	n.d.	-	n.d.	n.d.	
VF-10 (21-01-22)	C40_ET	F	J	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C41_ET	F	J	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C42_ET	M	J	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
VF-11 (21-01-29)	C43_ET	F	J	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C44_ET	M	J	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C45_ET	F	J	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C46_Ma	M	J	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
VF-13 (21-02-25)	C47_ET	F	J	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C48_ET	M	J	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C49_CSh	F	J	-	n.d.	n.d.	Pos.	LPAI	H5Nx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
VCa-1 (21-02-12)	C50_NP	A	M	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C51_NP	A	F	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
	C52_NP	J	F	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	
VCa-1 (21-02-15)	C53_NP	n.a.	n.a.	-	n.d.	n.d.	Pos.	LPAI	HxNx	-	n.d.	n.d.	-	n.d.	n.d.	-	n.d.	n.d.	

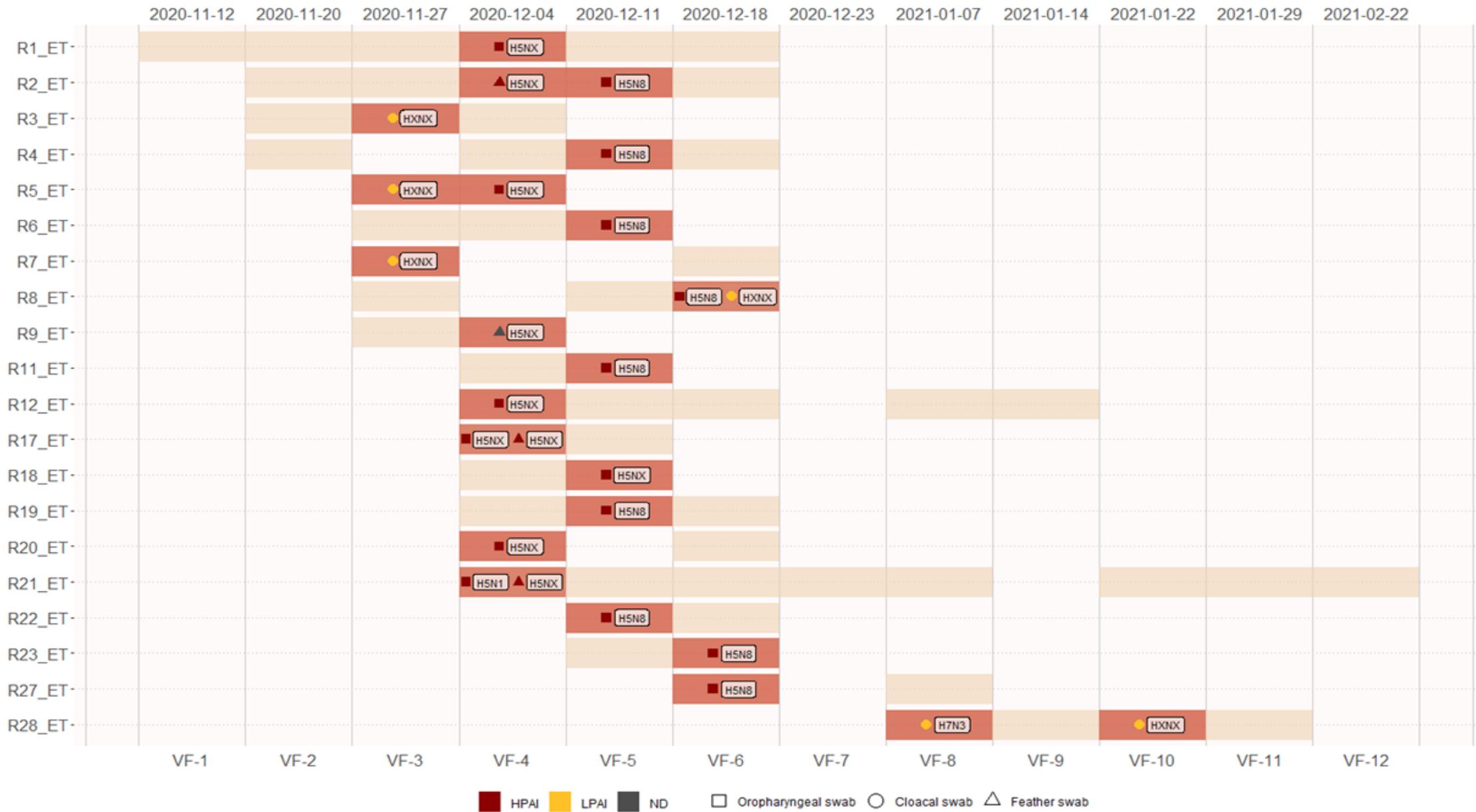
# HPAI epidemic curves in captured and hunted wild birds during late autumn-winter seasons 2020-2021 in Northern Est Italy



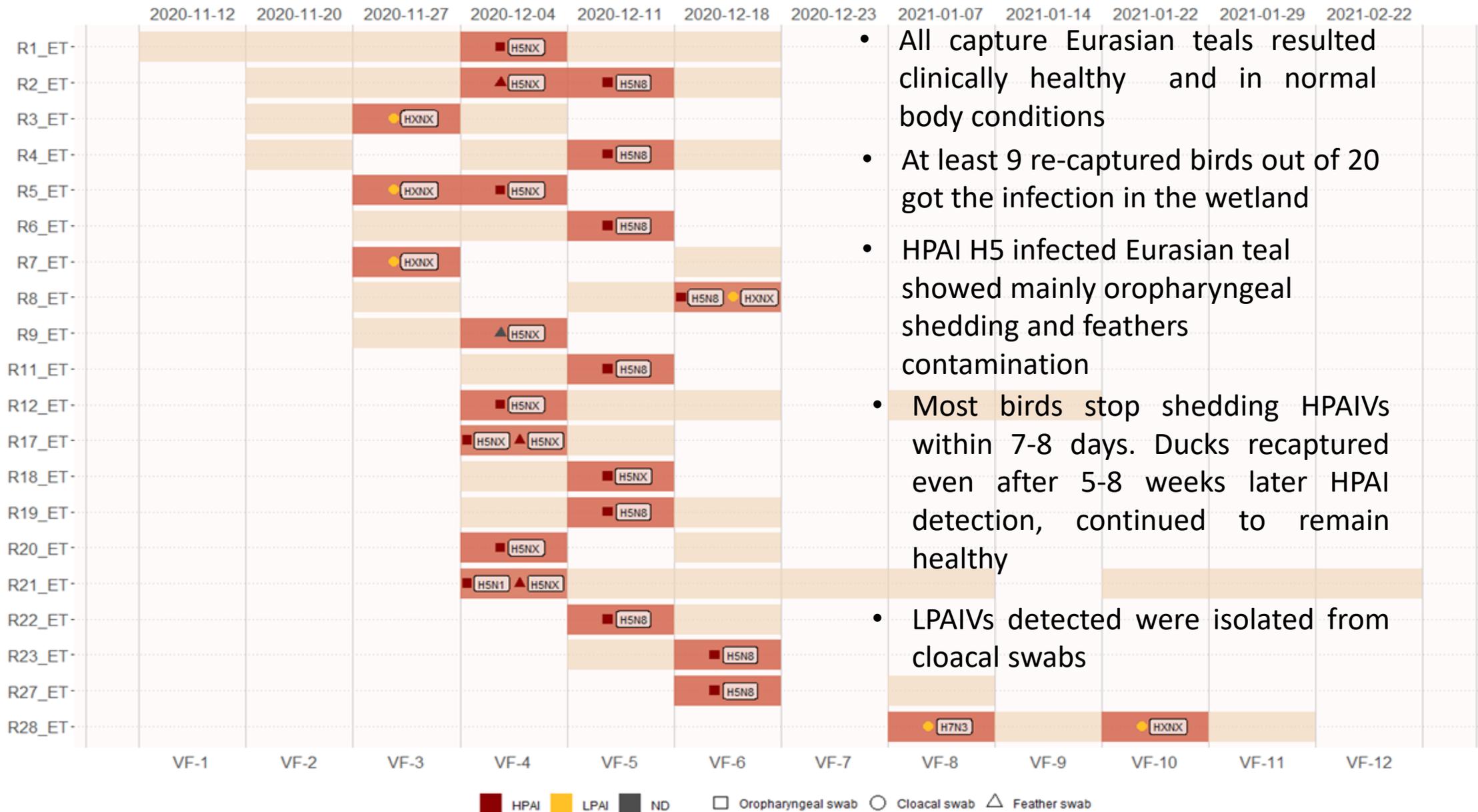
Capture station in Valle Figheri (Venice province)



# Progression of the HPAI H5 clade 2.3.4.4b infection in naturally infected wild migratory Eurasian teals



# Progression of the HPAI H5 clade 2.3.4.4b infection in naturally infected wild migratory Eurasian teals



- All capture Eurasian teals resulted clinically healthy and in normal body conditions
- At least 9 re-captured birds out of 20 got the infection in the wetland
- HPAI H5 infected Eurasian teal showed mainly oropharyngeal shedding and feathers contamination
- Most birds stop shedding HPAIVs within 7-8 days. Ducks recaptured even after 5-8 weeks later HPAI detection, continued to remain healthy
- LPAIVs detected were isolated from cloacal swabs

# Conclusions

- The results obtained in these studies are particular and reflect a different scenario to the one observed last winter in northern European countries
- Late autumn - early winter seems to be the period at higher risk of introduction of HPAIVs from migratory birds in southern European countries
- The early detection of HPAIV presence in wintering migratory wild birds must lead MS to the immediate implementation of adequate risk mitigation measures

# Conclusions

- Our findings confirm previous scientific evidences of Eurasian teal and Eurasian wigeon as long-distance vectors of HPAI H5Nx GsGd viruses 2.3.4.4b, since the infection may often occur asymptotically despite evident oropharyngeal and cloacal viral shedding
- Our data show that it is more probable to detect H5 HPAI clade 2.3.4.4b in oropharyngeal and feather swabs of dabbling ducks rather than in cloacal swabs.
- In order to detect more efficiently the early introduction of novel HPAI viruses, passive surveillance should be complemented by a targeted active surveillance tailored to each country

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## Active Surveillance for Highly Pathogenic Avian Influenza Viruses in Wintering Waterbirds in Northeast Italy, 2020–2021

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**Abstract:** The increasing involvement of wild waterfowl in H5 Highly Pathogenic Avian Influenza Virus (HPAIV) circulation continues to pose a threat to animal and public health worldwide. In winter 2020–2021, two field surveillance activities were carried out on a weekly basis, through virological and serological analyses, in 823 hunted and 521 trapped migratory aquatic birds in northeast Italy. Sixty Eurasian teals were recaptured several times, which allowed us to follow the progression of the HPAI H5 infection in naturally infected wild waterfowl. Oropharyngeal, cloacal, and feather swabs (CS, CS and FS) were collected from each duck and tested by real time rRT-PCR Type A influenza. The identified viruses were characterized and pathotyped by sequencing. Several viruses belonging to three different HPAI H5 subtypes were detected: H5N8, H5N5, and H5N1. High prevalence of infection with HPAI H5 clade 2.3.4.4b during November–December 2020 (up to 27.1%) was observed in captured Eurasian teals, while infection rates in hunted dabbling ducks, mainly Eurasian wigeons, showed the highest prevalence of infection in November 2020 (8.9%) and January 2021 (10.2%). All HPAI positive birds were also clinically healthy when recaptured weeks apart. The OS and FS showed the highest detection efficiency of HPAIV. Our results highlight that HPAI passive surveillance should be complemented by a targeted active surveillance to more efficiently detect novel HPAI viruses.

**Keywords:** avian influenza virus; HPAI H5 subtypes clade 2.3.4.4b; migratory aquatic bird; active surveillance



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